

HMM for sequence alignment:
profile HMM

Pair HMM

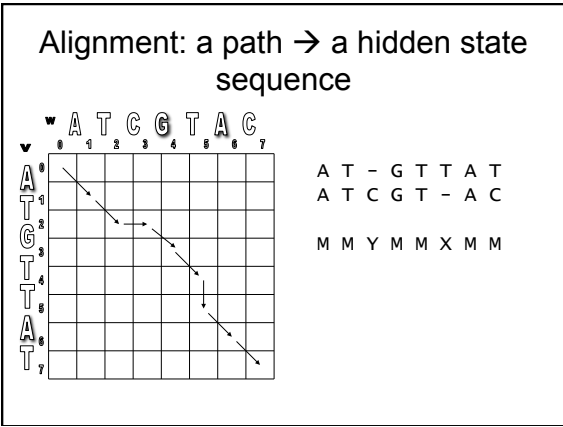
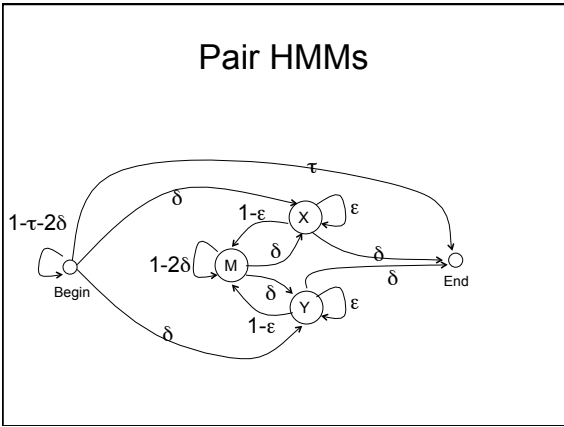
HMM for pairwise sequence alignment, which incorporates affine gap scores.

"Hidden" States

- Match (M)
- Insertion in x (X)
- insertion in y (Y)

Observation Symbols

- Match (M): $\{(a,b) \mid a,b \in \Sigma\}$.
- Insertion in x (X): $\{(a,-) \mid a \in \Sigma\}$.
- Insertion in y (Y): $\{(-,a) \mid a \in \Sigma\}$.



Multiple sequence alignment (Globin family)

```

Helix      AAAAAAAAAAAAAA  BBBBHHHHHHHHHHCCCCCCCC
HBA_HUMAN  -----VLEPAKTVKAAKQVCA--KAGCYGALSSNPLFFPTFTTFPF
HBB_HUMAN  -----VHLTEEKSAVTAWQVY---NVDEVGALGRLLVVPYQRTFFSFP
MYG_PHYCA  -----VLECGDQLVLRVWAKVEA--DVAGHQDILLKLFKHPETLEKDFDP
GLB1_CHITP -----LSEKQISYVQASFKVQG---DPIVLIYVYKADPSIKMPTQF
GLB5_PETMA PIVDVGVAFLSAARKETIESAMAPYS--TYETSVDLIVKFFSTPAAGFFPKF
LGB2_LUPLI -----GALTESQALVRESWKEFPA--NIKPTHTFFILVLEIAPAKDLFS-P
GLB1_GLYDI -----QLSAGKQVFAKWTWIKGAKGKQVQKCKLKEIQAHRQKAVYQ-P
Consensus  Ls... v a W kv . . g . L . f . P . F F
Helix      DDDDDDEEEEEEEEEEEEEEEEE  FFFFFFFF
HBA_HUMAN  -DLS---HUSAQVKGKGVADALTNAAVY--D--DMPNLDALSDJAHKIL-
HBB_HUMAN  GLETFPDAVKNVFAKGERVIGAPSSGLAH--D--NKQTFALISELJCKL-
MYG_PHYCA  KHLKTEAKWASEDKKQGVTVLTAIDALIKK---K-GHEAELEFLAQSHATKH-
GLB1_CHITP AG-KLESIKGTAFPTTHAKRIVGFFSKIGEL--P--NLEADVYTFVASHKPRG-
GLB5_PETMA KGLTADQLKSNADYRHWREIITMAVDVAEM--DPEKMGKELDLSGHRKSP-
LGB2_LUPLI LK-GTSKVPQNPPELQAHGKVFLLVYEAALQLQVTVVYDATALKNGSVHVKG-
GLB1_GLYDI SG---AS---DPGVAAIGAKVLAIGVAVSL--SDBGKVAQKAGVVRHKGYN
Consensus  t . . y . Hg kv . a a . l g . a . l . l H .
Helix      PFGCGCGCGCGCGCGCGCGGG  HHNNHHNNHHNNHHNNHHNNHH
HBA_HUMAN  -EVDPNFELLSCGLVFLAAHLPAEFTFAVAGLEKFLASVYVLSVYV---
HBB_HUMAN  -HYDEFNLLGQVLCVFLASRFGEFTFVQAVQKVAGVANALAEKTH----
MYG_PHYCA  -KIPKYLEIIEEALIVLHSHRDPFADGAGQAKNALLEPKDIAEYFELQVQ-
GLB1_CHITP --VTDQNLNFRFAGVSYWMAAT--DFA-QAQAANQVLEFFQVFM-----
GLB5_PETMA -QVDPQVFLAAVADVVAAG-----DAGFEKMMMCILLRSAY-----
LGB2_LUPLI --YAKRFVYEAALKTIEEYVAKWSEELNSWTLATIEALVLIKRENDAA--
GLB1_GLYDI KHIAQTFEFLGASLLSMEHRIGGQNAAKDAAAYADISGLISGSGS----
Consensus  v . f l . . . . . f . aa . k . . l sky
  
```

Profile model (PSSM)

- A natural probabilistic model for a conserved region would be to specify independent probabilities $e_i(a)$ of observing nucleotide (amino acid) a in position i
- The probability of a new sequence x according to this model is

$$P(x|M) = \prod_{i=1}^L e_i(x_i)$$

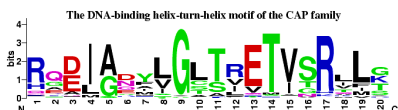
Profile / PSSM

•DNA / proteins Segments of the same length L;

•Often represented as Positional frequency matrix;

```

LTMRGDIIGNYLGLTETISRLLGRFQKSGML
LTMRGDIIGNYLGLTETISRLLGRFQKSGMI
LTMRGDIIGNYLGLTETISRLLGRFQKSEIL
LTMRGDIIGNYLGLTETISRLLGRQKMGIL
LAMSREIIGNYLGLAVETVSRVFSRFQNGELI
LAMSREIIGNYLGLAVETVSRVTRFQQNGLI
LPMSRNEIIGNYLGLAVETVSRVTRFQQNGLL
VRMSREEIIGNYLGLTETVSRVLSRFQREGLI
LRMSREEIIGNYLGLKLETVSRVLSRFQREGLI
LPMCRDIDYDLGLTETVSRVLSQLHTQGIL
LPMSRRDIADYDLGLTETVSRVLSQLHTDGLV
LPMSRQDIADYDLGLTETVSRVTKLERHGAI
    
```



Searching profiles: inference

• Give a sequence S of length L, compute the likelihood ratio of being generated from this profile vs. from background model:

$$R(S|P) = \frac{\prod_{i=1}^L e_i(x_i)}{q_{x_i}}$$

– Searching motifs in a sequence: sliding window approach

Match states for profile HMMs

• Match states

– Emission probabilities $e_{M_i}(a)$



Components of profile HMMs

• Insert states $e_i(a)$

– Emission prob.

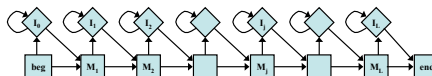
• Can be the background distribution q_a .

– Transition prob.

• M_i to I_i , I_i to itself, I_i to M_{i+1}

– Log-score for a gap of length k (not including the log-score from emission)

$$\log a_{M_i I_i} + \log a_{I_i M_{i+1}} + (k-1) \log a_{I_i I_i}$$



Components of profile HMMs

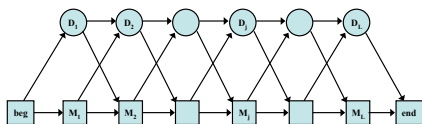
• Delete states

– No emission prob.

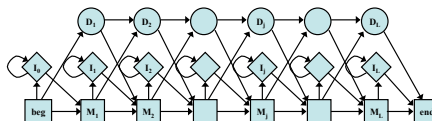
– Cost of a deletion

• M_i to D_{i-1} , D_i to D_{i+1} , D_i to M_{i+1}

• Each D_i to D_{i+1} might be different for different i



Full structure of profile HMMs



This is the structure implemented in Hmmer, slightly different from the structure described in the textbook: there is no transition allowed from D_i to I_i or from I_i to D_{i+1} . As a result, the recursive equation for Viterbi algorithm is different from the one described in the book too.

Deriving HMMs from multiple alignments

- Key idea behind profile HMMs
 - Model representing the consensus for the alignment of sequence from the same family
 - Not the sequence of any particular member

```

HBA_HUMAN  ...VGA--HAGEY...
HBB_HUMAN  ...V----NVDEV...
MYG_PHYCA  ...VEA--DVAGH...
GLB3_CHITP  ...VKG-----D...
GLB5_PETMA  ...VYS--TYETS...
LGB2_LUPLU  ...FNA--NIPKH...
GLB1_GLYDI  ...IAGADNGAGV...
*** *****
    
```

Deriving HMMs from multiple alignments

- Basic profile HMM parameterization
 - Aim: making the higher probability for sequences from the family
- Parameters
 - the transition and emission probabilities: trivial if many of independent alignment sequences are given.

$$a_{kl} = \frac{A_{kl}}{\sum_{l'} A_{kl'}} \quad e_k(a) = \frac{E_k(a)}{\sum_{a'} E_k(a')}$$

- length of the model: heuristics or systematic way (e.g., using the MAP algorithm)

Deriving HMMs from multiple alignments

(a) Multiple alignment:

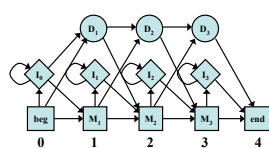
```

      x x . . . x
bat   A G - - - C
rat   A G A G - C
cat   A G - A A G
gnat  - G A A A C
goat  A G - - - C
Matching 1 2 . . . 3
    
```

(c) Observed emission/transition counts

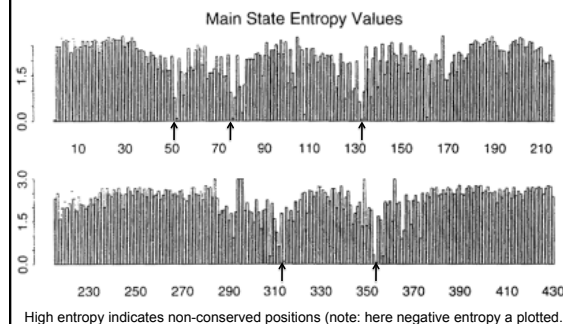
Emission from M	A	0	1	2	3
C	-	4	0	0	0
G	-	0	5	1	0
T	-	0	0	0	0
Emission from I	A	0	0	6	0
C	0	0	0	0	0
G	0	0	0	1	0
T	0	0	0	0	0
Transition probabilities	M-M	4	4	2	4
M-D	1	0	0	0	
M-I	0	0	3	0	
I-M	0	0	2	0	
I-I	0	0	4	0	
D-M	-	1	0	0	
D-D	-	0	0	0	

(b) Profile-HMM architecture:



A simple rule that works well is that columns that are more than half gap characters should be modeled by inserts.

Sequence conservation: entropy of the emission probability distributions



High entropy indicates non-conserved positions (note: here negative entropy is plotted).

Matching a sequence to a profile HMM (global alignment)

- Viterbi algorithm: pHMM θ (with L matching states) and a query sequence $x_1 x_2 \dots x_N$

$$V_j^M(i) = e_{M_j}(x_i) \max \begin{cases} V_{j-1}^M(i-1) \cdot a_{M_{j-1}M_j} \\ V_{j-1}^I(i-1) \cdot a_{I_{j-1}M_j} \\ V_{j-1}^D(i-1) \cdot a_{D_{j-1}M_j} \end{cases}$$

$$V_j^I(i) = e_{I_j}(x_i) \max \begin{cases} V_j^M(i-1) \cdot a_{M_j I_j} \\ V_j^I(i-1) \cdot a_{I_j I_j} \end{cases}$$

$$V_j^D(i) = \max \begin{cases} V_{j-1}^M(i) \cdot a_{M_{j-1}D_j} \\ V_{j-1}^I(i) \cdot a_{D_{j-1}D_j} \end{cases}$$

for $i=1, \dots, L, j=1, \dots, N$

Note: this is slightly different from the textbook; no transition from D_j to I_j or from I_j to D_{j+1} .

Viterbi algorithm: trace back

```

PrintStateSeq(ptr^M, ptr^I, ptr^D, ptr^N, i, j)
if i=0 or j=0
    return;
if ptr^M(L) = "M"
    printStateSeq(ptr^M, ptr^I, ptr^D, "M", i-1, j-1)
else if ptr^M(L) = "I"
    printStateSeq(ptr^M, ptr^I, ptr^D, "I", i-1, j-1)
else if ptr^M(L) = "D"
    printStateSeq(ptr^M, ptr^I, ptr^D, "D", i-1, j-1)
print M
else if ptr^M(L) = "I"
    printStateSeq(ptr^M, ptr^I, ptr^D, "M", i, j-1)
else if ptr^M(L) = "D"
    printStateSeq(ptr^M, ptr^I, ptr^D, "I", i, j-1)
print I
else
    if ptr^M(L) = "I"
        printStateSeq(ptr^M, ptr^I, ptr^D, "M", i, j-1)
    else if ptr^M(L) = "D"
        printStateSeq(ptr^M, ptr^I, ptr^D, "D", i, j-1)
    print D
PrintStateSeq(ptr^M, ptr^I, ptr^D, ptr^N, N, L)
    
```

Matching a sequence to a profile HMM (global alignment)

- Forward: pHMM θ (with L matching states) and a query sequence $x_1 x_2 \dots x_N$

$$F_j^M(i) = e_{M_j}(x_i) \cdot [F_{j-1}^M(i-1) \cdot a_{M_{j-1}M_j} + F_{j-1}^I(i-1) \cdot a_{I_{j-1}M_j} + F_{j-1}^D(i-1) \cdot a_{D_{j-1}M_j}]$$

$$F_j^I(i) = e_{I_j}(x_i) \cdot [F_j^M(i-1) \cdot a_{M_j I_j} + F_j^I(i-1) \cdot a_{I_j I_j}] \quad \text{for } i=1, \dots, L, j=1, \dots, N$$

$$F_j^D(i) = F_{j-1}^M(i) \cdot a_{M_{j-1}D_j} + F_{j-1}^I(i) \cdot a_{D_{j-1}D_j}$$

Initialization: $F_0^M(0) = 1; F_{j>0}^M(0) = 0; F_0^M(i > 0) = 0; F_j^I(0) = 0; F_0^D(i) = 0;$

Termination: $F = F_L^M(N) + F_L^I(N) + F_L^D(N)$

Note: this is slightly different from the textbook; no transition from D_j to I_j or from I_j to D_{j+1} .

Example

Query: AGG (N=3)

		0	1	2	3
Emission from M	A	-	1	.5	0
	C	-	0	0	.8
	G	-	0	.5	.2
	T	-	0	0	0
Emission from I	A	.2	.2	.9	.2
	C	.3	.3	0	.3
	G	.3	.3	.1	.3
	T	.2	.2	0	.2
Transition probabilities	M-M	.8	1	.4	1
	M-D	.2	0	0	0
	M-I	0	0	.6	0
	I-M	.5	.5	.3	.5
I-I	.5	.5	.7	.5	
D-M	-	1	.5	1	
D-D	-	0	.5	0	

Example: Viterbi algorithm

Query: X=AGG (N=3)

Example: Viterbi algorithm

Query: X=AGG (N=3)

$$V_1^D(0) = \max(V_0^M(0) a_{M_0 D_1}, V_0^I(0) a_{I_0 D_1}) = \max(0, 0) = 0.2$$

Example: Viterbi algorithm

Query: X=AGG (N=3)

$$V_1^M(1) = e_{M_1}(x_1) \max(V_0^M(0) a_{M_0 M_1}, V_0^I(0) a_{I_0 M_1}, V_0^D(0) a_{D_0 M_1}) = 1 \times \max(1 \times 0.8 \times 0.5 \times 0.1) = 0.8$$

Example: Viterbi algorithm

Query: X=AGG (N=3)

$$V_3^M(3) = ?$$

Example: Viterbi algorithm

		0 1 2 3			
Emission from M	A	-1	.5	0	
	C	0	0	.8	
	G	0	.5	.2	
	T	0	0	0	
Emission from I	A	.2	.2	.9	.2
	C	.3	.3	0	.3
	G	.3	.3	.1	.3
	T	.2	.2	0	.2
Transition probabilities	M→M	.8	1	.4	1
	M→D	.2	0	0	0
	M→I	0	0	.6	0
	I→M	.5	.5	.3	.5
I→I	.5	.5	.7	.5	
D→M	0	1	.5	1	
D→D	0	0	.5	0	

		i 0 1 2 3			
0	V ^M	1	0	0	0
	V ^I	0	0	0	0
	V ^D	0	0	0	0
	V ⁰	0	.8	0	0
1	V ^M	0	.8	0	0
	V ^I	0	0	0	0
	V ^D	0	.2	0	0
	V ⁰	0	.1	.4	0
2	V ^M	0	0	.008	.0240
	V ^I	0	0	0	0
	V ^D	0	0	0	0
	V ⁰	0	0	0	0
3	V ^M	0	0	.008	0.320
	V ^I	0	0	0	.0014
	V ^D	0	0	0	0
	V ⁰	0	0	0	0

Traceback

Query: X=AGG (N=3)

Searching with profile HMMs

- Main usage of profile HMMs
 - Detecting potential sequences in a family
 - Core algorithm: matching a sequence to a profile HMMs
 - Viterbi algorithm or forward algorithm
 - Comparing the resulting probability with random model (R): log-odd score

$$P(x | R) = \prod_i q_{x_i}$$

where q_x is the frequency of observing x_i .

Matching a sequence to a profile HMM (global alignment)

Viterbi algorithm

$$V_j^M(i) = \log \frac{e_{M_j}(x_i)}{q_{x_i}} + \max \begin{cases} \log V_{j-1}^M(i-1) + \log a_{M_j, M_j}, & \text{Initialization:} \\ \log V_{j-1}^I(i-1) + \log a_{I_j, M_j}, & V_j^M(0) = 0; \quad V_{j+1}^M(0) = -\infty; \quad V_0^M(i > 0) = -\infty \\ \log V_{j-1}^D(i-1) + \log a_{D_j, M_j}; & V_j^I(0) = -\infty; \end{cases}$$

$$V_j^I(i) = \log \frac{e_{I_j}(x_i)}{q_{x_i}} + \max \begin{cases} \log V_{j-1}^M(i-1) + \log a_{M_j, I_j}, & V_0^I(i) = -\infty \\ \log V_{j-1}^I(i-1) + \log a_{I_j, I_j}; \end{cases}$$

$$V_j^D(i) = \max \begin{cases} \log V_{j-1}^M(i) + \log a_{M_j, D_j}, & \text{Termination:} \\ \log V_{j-1}^D(i) + \log a_{D_j, D_j}; & V = \max[V_i^M(N), V_i^I(N), V_i^D(N)] \end{cases}$$

for $i=1, \dots, L, j=1, \dots, N$

Matching a sequence to a profile HMM (global alignment)

Forward algorithm

$$F_j^M(i) = \log \frac{e_{M_j}(x_i)}{q_{x_i}} + \log[a_{M_j, M_j} \exp(F_{j-1}^M(i-1)) + a_{I_j, M_j} \exp(F_{j-1}^I(i-1)) + a_{D_j, M_j} \exp(F_{j-1}^D(i-1))];$$

$$F_j^I(i) = \log \frac{e_{I_j}(x_i)}{q_{x_i}} + \log[a_{M_j, I_j} \exp(F_j^M(i-1)) + a_{I_j, I_j} \exp(F_j^I(i-1))];$$

$$F_j^D(i) = \log[a_{M_j, D_j} \exp(F_j^M(i)) + a_{I_j, D_j} \exp(F_j^I(i))];$$

Initialization: $V_0^M(0) = 0; \quad V_{j+1}^M(0) = -\infty; \quad V_0^M(i > 0) = -\infty;$ Termination: $F = \log[\exp(F_N^M(N)) + \exp(F_N^I(N)) + \exp(F_N^D(N))]$
 $V_0^I(0) = -\infty;$
 $V_0^D(i) = -\infty.$

Significance of HMM alignment

- The log-odd score of local Viterbi alignment (V) alignment between a random sequence and a profile HMM follows a Gumbel (type I EVD) distribution

$$P(V \geq t) = 1 - \exp[-e^{-\lambda(t-\mu)}]$$
- With ~200 Viterbi, the location parameter μ can be accurately estimated;
- $\lambda \sim \log(z)$, z is the base of the log-odd score, e.g., $z=2$ when the sequence length approaches infinite
- The length effect can be corrected by $\lambda \sim \log 2 + \frac{1.44}{hN}$
 - Where, N is the length, and h is the average relative entropy per match state in the pHMM;
 - For typical Pfam models, $N \sim 140, h \sim 1.8, \lambda \sim \log 2 + 0.0057$, a small correction.

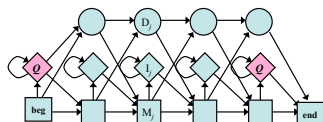
Eddy, PLoS Comp. Biol., 4:1, 2008

Variants for non-global alignments

- Local alignment (Smith-Waterman type)
 - Emission prob. in flanking states use background values q_x .
 - Looping prob. close to 1, e.g. $(1 - \eta)$ for some small η .

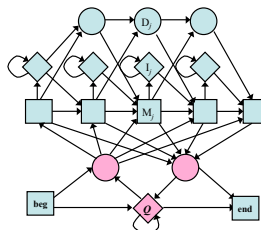
Variants for non-global alignments

- Overlap (also called *glocal* or *fit*) alignment
 - The loop probability of the *first* and *last* insert states is much higher than the other insert states
 - When expecting to find either present as a whole or absent (e.g., of a protein domain within a protein)
 - Transition to first delete state allows missing first residue



Variants for non-global alignments

- Repeat alignments
 - Transition from right flanking state back to random model
 - Can find multiple matching segments in query string



Optimal model construction: different ways of marking columns

(a) Multiple alignment:

```

bat      x x . . . x
rat      A - A G - - C
cat      A G - A A -
gnat     - - A A A C
goat     A G - - - C
Matching 1 2 . . . 3
    
```

(b) Profile-HMM architecture:

(c) Observed emission/transition counts

		0	1	2	3
Emission from M	A	-	4	0	0
	C	-	0	0	4
	G	-	0	3	0
	T	-	0	0	0
Emission from I	A	0	0	6	0
	C	0	0	0	0
	G	0	0	1	0
	T	0	0	0	0
Transition probabilities	M-M	4	3	2	4
	M-D	1	1	0	0
	M-I	0	0	1	0
	I-M	0	0	2	0
	I-D	0	0	1	0
	I-I	0	0	4	0
	D-M	-	0	0	1
	D-D	-	1	0	0
D-I	-	0	2	0	

Optimal model construction

- MAP (match-insert assignment)
 - Recursive calculation of a number S_j
 - S_j : log prob. of the optimal model for alignment up to and including column j , assuming j is marked.
 - S_j is calculated from S_i and summed log prob. between i and j .
 - T_{ij} : summed log prob. of all the state transitions between marked i and j .
- $$T_{ij} = \sum_{x,y \in \{M,D,I\}} c_{xy} \log a_{xy}$$
- c_{xy} are obtained from partial state paths implied by marking i and j .

Optimal model construction

- Algorithm: MAP model construction
 - Initialization:
 - $S_0 = 0, M_{L+1} = 0$.
 - Recurrence: for $j = 1, \dots, L+1$:

$$S_j = \max_{0 \leq i < j} (S_i + T_{ij} + M_j + I_{i+1,j-1} + \lambda)$$

$$\sigma_j = \arg \max_{0 \leq i < j} (S_i + T_{ij} + M_j + I_{i+1,j-1} + \lambda)$$
 - Traceback: from $j = \sigma_{L+1}$, while $\sigma_j > 0$:
 - Mark column j as a match column
 - $j \leftarrow \sigma_j$.

Weighting training sequences

- Input sequences are random?
- “Assumption: all examples are independent samples” might be incorrect
- Solutions
 - Weight sequences based on similarity: highly similar pair of training sequences receive lower weights

Multiple sequence alignment (MSA) by training profile HMM

- Sequence profiles can be represented as probabilistic models like profile HMMs.
 - ML methods for building (training) profile HMM are based on multiple sequence alignment
 - Profile HMMs can also be trained from initially unaligned sequences using the Baum-Welch-like EM algorithm
 - Simultaneously aligning multiple sequences and building the profile HMM from the multiple alignment

Multiple alignment with a known profile HMM

- A step backward: to derive a multiple alignment from a known profile HMM model
 - e.g., to align many sequences from the same family based on the HMM model built from the (seed) multiple alignment of a small representative set of sequences in the family.
- It just requires calculating a Viterbi alignment for each individual sequence
 - Match a sequence to a profile HMM: Viterbi algorithm
 - Residues aligned to the same match state in the profile HMM should be aligned in the same columns;
 - Given a preliminary alignment, HMM can align additional sequences.

Multiple alignment with a known profile HMM

- Comparing with other MSA program
 - Profile HMM does not align inserts whereas other MSA algorithms align the whole sequences.

• Position	1	2	3	4	5	6	insert	7	8	9	10	11	
	F	P	H	F	-	D	LS		H	G	S	A	Q
	F	E	S	F	G	D	LSTPDAV	M	G	N	P	K	
	F	D	R	F	K	H	LKTEAEM	K	A	S	E	D	
	F	T	Q	F	A	G	KDLEST	K	G	T	A	P	
	F	P	K	F	K	G	LTADQL	K	K	S	A	D	
	F	S	-	F	L	K	GTSEVP	Q	N	N	P	E	
	F	G	-	F	S	G	AS	-	-	D	P	G	

Training profile HMM from unaligned sequences

- Simultaneously aligning multiple sequences and building the profile HMM from the multiple alignment
 - Initialization: choose the length of the profile HMM and initialize parameters of the model
 - MSA: align all sequences to the final model using the Viterbi algorithm and build a multiple alignment as described in the previous section.
 - Training: estimate the model using the Baum-Welch algorithm
 - Iterating until the model (and the MSA) converges

Profile HMM training from unaligned sequences

- Initial Model
 - The only decision that must be made in choosing an initial structure for Baum-Welch estimation is the length of the model M.
 - A commonly used rule is to set M be the average length of the training sequence.
 - We need some randomness in initial parameters to avoid local maxima.

Multiple alignment by profile HMM training

- Avoiding Local maxima
 - Baum-Welch algorithm is guaranteed to find a LOCAL maxima.
 - Models are usually quite long and there are many opportunities to get stuck in a wrong solution.
 - Solution
 - Start many times from different initial models.
 - Use some form of stochastic search algorithm, e.g. simulated annealing.

Multiple alignment by profile HMM training--Model surgery

- We can modify the model after (or during) training a model by manually checking the alignment produced from the model.
 - Some of the match states are redundant
 - Some insert states absorb too many sequences
- Model surgery
 - If a match state is used by less than 1/2 of training sequences, delete its module (match-insert-delete states)
 - If more than 1/2 of training sequences use a certain insert state, expand it into n new modules, where n is the average length of insertions
 - ad hoc, but works well

Hmmer 3

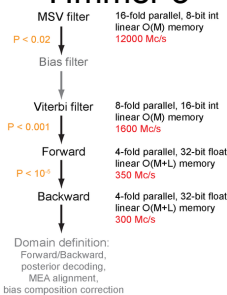


Figure 4. The HMMER3 acceleration pipeline.

Eddy SR (2011) Accelerated Profile HMM Searches. PLoS Comput Biol 7(10): e1002195. doi:10.1371/journal.pcbi.1002195



Accelerated Profile HMM Searches

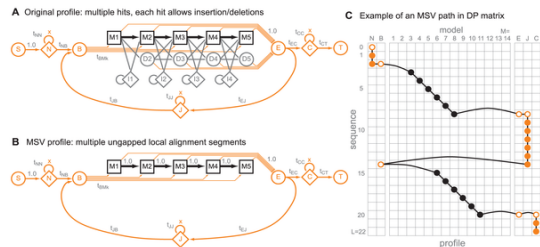


Figure 1. The MSV profile.

Eddy SR (2011) Accelerated Profile HMM Searches. PLoS Comput Biol 7(10): e1002195. doi:10.1371/journal.pcbi.1002195

